



Diagnostic Exercise
From The Latin Comparative Pathology Group*

Clostridial enterocolitis in an emu (*Dromaius novaehollandiae*)

Contributors:

Emily Hoskins¹, Lauren W. Stranahan¹, Alice Blue-McLendon^{1,2}, Raquel Rech^{1*}

¹ Department of Veterinary Pathobiology

²Department of Veterinary Physiology and Pharmacology,

College of Veterinary Medicine and Biomedical Sciences, Texas A&M University, College Station, TX, USA

**Corresponding author: E-mail: rrech@cvm.tamu.edu

Clinical History:

A 25-year-old female emu (*Dromaius novaehollandiae*) presented with a 1-week history of lethargy, hyporexia, and recumbency. The bird had lived its entire life at the Winnie Carter Wildlife Center in Texas and had an unremarkable medical history, apart from bilateral congenital blindness.

Necropsy Findings:

Within the coelomic cavity, the serosal surface of the intestines was coated with a moderate amount of tan to green, mucoid, opaque, foul-smelling fluid (fibrinoheterophilic coelomitis) (Fig. 1).

The large intestine was diffusely dilated and filled with dark red fluid and strands of yellow, friable material (fibrin). The mucosa of the small intestine, and most severely the colon, was diffusely roughened, thickened, dull, and covered by a thick yellow to green, opaque, friable diphtheritic membrane (Fig. 2). The distal colon was diffusely, markedly distended and focally contained a 50x7x5 cm, semi-firm accumulation of feces coated by a 4-5 mm diameter layer of tan, friable material (fibrin) located 21 cm from the opening of the cloaca. Gross evaluation of the eyes revealed bilateral aphakia.

Follow-up questions:

- Morphologic diagnosis
- Ancillary tests to determine the etiology

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<https://davisthompsonfoundation.org/diagnostic-exercise/>

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Gross and Microscopic Images:



Figure 1. The colon is markedly distended and covered with multifocal strands of fibrin



Figure 2. A thick diphtheritic membrane composed of fibrin and necrotic debris coats the small intestinal mucosa.

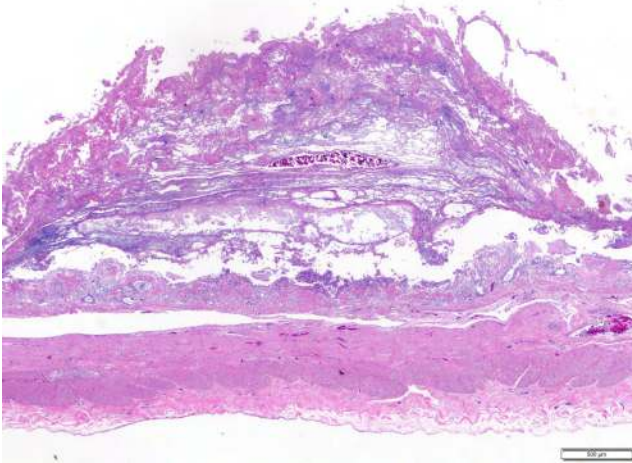


Figure 3. The small intestinal mucosa is effaced by a thick membrane composed of necrotic cellular debris and fibrin. Ha&E stain.

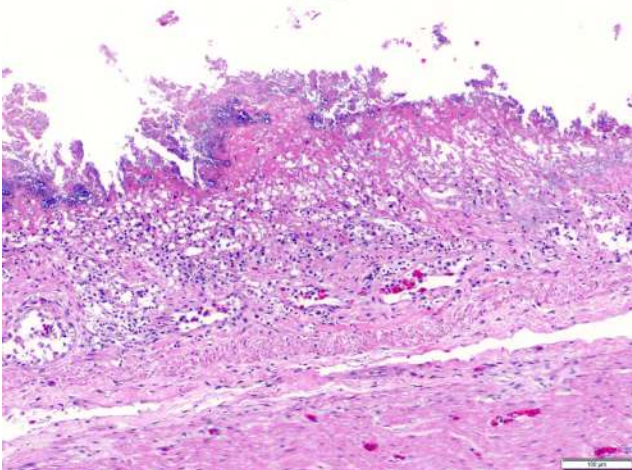


Figure 4. Large colonies of 1x2 µm bacilli coat the surface of the mucosa of the intestine. H&E stain.

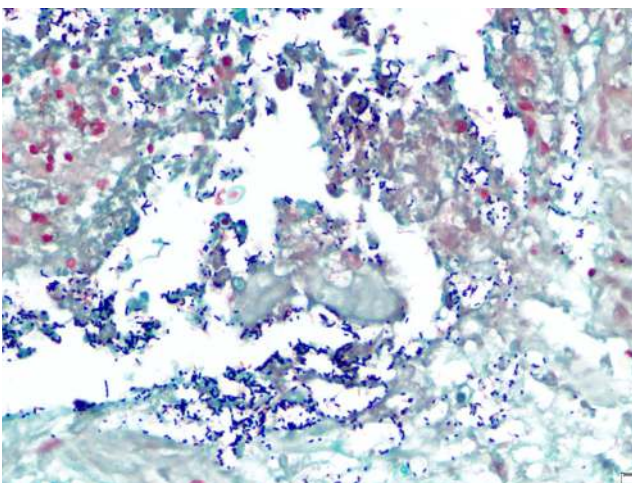


Figure 5. Gram-positive bacilli on the mucosal surface of the intestine. Gram stain.

- **Morphologic diagnosis:** Marked, diffuse, acute, fibrinonecrotizing enterocolitis with myriad bacterial cocci.
- **Ancillary test results:**
 - PCR for toxins A and B of *Clostridioides difficile* on the small intestinal contents was positive
 - PCR for *Salmonella* spp. on intestinal contents was negative
 - Gram stain: numerous gram-positive bacilli coating the mucosa
- **Etiology:** *Clostridioides difficile*

Comments:

Clostridioides difficile is a gram-positive, anaerobic bacillus that can cause colitis-associated diarrhea in humans and many other mammals including hamsters, guinea pigs, rabbits, horses, and neonatal pigs (4). Gastrointestinal lesions associated with a *C. difficile* infection can vary greatly in location and severity depending on the species, age, bacterial spore presence, and the individual animal response to disease. In general, necropsy findings include an aborally distributed colitis, typhlitis, and occasionally ileitis (4). Lesions in hamsters and guinea pigs most commonly include a hemorrhagic typhlitis that range from petechiation to diffuse hemorrhagic lesions in the cecum, colon, and ileum (1, 5). Several reports describe chronic typhlitis associated with mucosal hyperplasia in addition to cholangiohepatitis and hepatic, renal, and intestinal amyloidosis (1, 2). Infections in the pig typically lead to a mild typhlocolitis that can sometimes progress to transmural necrosis with microscopic, fibrin and neutrophil-secreting ulcers known as “volcano ulcers.” The cecum and colon are often distended by edema. Pigs can clinically present with diarrhea, obstipation, or dyspnea associated with ascites or hydrothorax (4). Severe cases in adult horses can cause a hemorrhagic necrotizing typhlocolitis (4) while in foals and rabbits, lesions are typically seen in the small intestine (6, 7). Various risk factors for development of *C. difficile* infection in animals have been identified, and include antimicrobial treatment, change in diet, and environmental stressors such as transportation (4). No known changes in this bird’s routine occurred prior to development of enterocolitis and so the predisposing factors in this case are uncertain. Although *C. difficile* infections are less common in avian species, there have been several reports of this disease in captive ostriches (3) in addition to *C. difficile*-associated hepatitis in ostrich chicks (8). It has also been isolated from the intestine of ostriches, rheas, and emus (10) and should be considered as a differential diagnosis when ratites present with enterocolitis. While an abundance

of clostridial growth on culture of the intestine allows for a presumptive diagnosis, ELISA or PCR on intestinal contents in order to detect clostridial toxins A and B (9) is recommended for definitive diagnosis, as in this case.

References

1. Bartlett JG, Chang TW, Moon N, Onderdonk AM. Antibiotic-induced lethal enterocolitis in hamsters: studies with eleven agents and evidence to support the pathogenic role of toxin-producing *Clostridia*. Am J Vet Res. 1978;39(9):1525-30.
2. Chang J, Rohwer RG. *Clostridium difficile* infection in adult hamsters. Lab Anim Sci. 1991;41(6):548-52.
3. Frazier KS, Herron AJ, Hines II ME, Gaskin JM, Altman NH. Diagnosis of enteritis and enterotoxemia due to *Clostridium difficile* in captive ostriches (*Struthio camelus*). J Vet Diagn Invest. 1993;5(4):623-5.
4. Keel MK, Songer JG. The comparative pathology of *Clostridium difficile*-associated disease. Vet Pathol. 2006;43(3):225-40.
5. Lowe BR, Fox JG, Barlett JG. *Clostridium difficile*-associated cecitis in guinea pigs exposed to penicillin. Am J Vet Res. 1980;41(8):1277-9.
6. Magedesian KG, Hirsh DC, Jang SS, Hansen LM, Madigan JE. Characterization of *Clostridium difficile* isolates from foals with diarrhea: 28 cases (1993-1997). J Am Vet Assoc. 2002;220(1):67-73.
7. Perkins SE, Fox JG, Taylor NS, Green DL, Lipman NS. Detection of *Clostridium difficile* toxins from the small intestine and cecum of rabbits with naturally acquired enterotoxemia. Lab Anim Sci. 1995;45(4):379.
8. Shivaprasad HL. Hepatitis associated with *Clostridium difficile* in an ostrich chick. Avian Pathol. 2003;32(1):57-62.
9. Sullivan NM, Pellett S, Wilkins TD. Purification and characterization of toxins A and B of *Clostridium difficile*. Infect Immun. 1982;35(3):1032-40.
10. Terio KA, McAloose D, Mitchell (née Lane) E. Palaeognathae: Apterygiformes, Casuariiformes, Rheiformes, Struthioniformes; Tinamiformes. In: Pathology of Wildlife and Zoo Animals. Ames: Elsevier; 2018. P. 263-85.