







Case Report

Transmural gastrointestinal stromal tumor of the mixed subtype in a canine: case report

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Abstract

Gastrointestinal stromal tumors (GISTs) are mesenchymal neoplasms originating from interstitial cells of Cajal, characterized, in most cases, by the immunohistochemical expression of c-

27 Kit (CD117). In dogs, they are rare and the associated clinical signs are nonspecific, including
28 vomiting, diarrhea, weight loss, and abdominal pain, which often makes clinical diagnosis difficult.
29 This paper describes a case of GIST in a dog, located in the cecum, detected through imaging exams
30 and diagnosed via anatomopathological evaluation, and immunohistochemical. Macroscopically, the
31 mass was transmural and measured approximately 7.0 cm in diameter, white to dark brown in color,
32 had a soft consistency, and, upon cutting, a smooth, solid surface. Microscopically, the neoplasm was
33 predominantly composed of spindle cells, with multifocal areas of epithelioid cells, both with positive
34 plasma membrane staining for c-Kit and absence of Desmin expression, confirming the diagnosis.
35 This report reinforces the importance of immunohistochemical evaluation in differentiating GISTs
36 from other mesenchymal neoplasms of the gastrointestinal tract and contributes to the understanding
37 of the morphological and clinicopathological aspects of this entity in dogs.

38

39 **Keywords:** c-Kit, dog, GIST, immunohistochemistry, mesenchymal neoplasm.

40

41 **Introduction**

42

43 Gastrointestinal neoplasms in dogs are considered rare, representing less than 3% of tumors
44 affecting the species (12). Despite their infrequency, a wide variety of diagnoses can be observed
45 within this group, with distinct histogenesis (3, 8, 9, 13, 16). Among gastrointestinal neoplasms, the
46 most common diagnoses are adenocarcinomas, lymphomas, leiomyosarcomas, and gastrointestinal
47 stromal tumors (GISTs) (22).

48 GISTs are mesenchymal neoplasms found primarily in the intestine and stomach (4). They are
49 believed to originate from the interstitial cells of Cajal, known as the pacemaker of the gastrointestinal
50 system and responsible for regulating intestinal motility (16, 19).

51 Recent studies highlight a higher occurrence in mixed-breed dogs, Dachshunds, Poodles,
52 Labradors, and Golden Retrievers (1, 4). There is no clear gender predilection, although there are

53 reports indicating a higher number of cases in males (1, 4, 8). Middle-aged and older animals are the
54 most affected, with a higher prevalence in dogs over 8 years old (1, 4, 8, 9).

55 Because of their histopathological characteristics similar to smooth muscle neoplasms, it is
56 estimated that, in the past, a significant portion of leiomyosarcoma diagnoses in dogs actually
57 corresponded to GISTs (16, 19). Differentiation from other neoplasms is mainly made by the distinct
58 immunohistochemical profile of GISTs. In this context, Cajal cells express positive staining for c-Kit
59 proteins, which are transmembrane tyrosine kinase receptor proteins involved in cell differentiation
60 and proliferation, and DOG1, a calcium-dependent chloride channel. Leiomyosarcomas have positive
61 expression for desmin, a widely used smooth muscle marker, while Cajal cells from GISTs generally
62 do not express this protein (4, 5, 17).

63 Therefore, specific diagnostic determination is fundamental for the appropriate therapeutic
64 choice, since GISTs are responsive to therapy with tyrosine kinase inhibitors, which does not occur
65 with other non-hematopoietic sarcomas (4). Thus, the objective of this work is to report a case of
66 gastrointestinal stromal tumor in a dog, highlighting the histopathological and immunohistochemical
67 aspects.

68

69 **Case description**

70

71 A 12-year-old female mixed-breed dog was treated at the University Veterinary Medicine
72 Hospital of the Federal Fluminense University with a chief complaint of a nodule in the ventral
73 cervical region, with a suspected diagnosis of recurrence of cutaneous mast cell tumor. The animal
74 presented with obesity, pale mucous membranes, abdominal tenderness, enlarged prescapular and
75 popliteal lymph nodes, and a grade I/IV mitral valve murmur.

76 An abdominal ultrasound examination was performed, which revealed a mass of approximately
77 7.0 centimeters in the mesogastric region, near the transverse colon. Cytopathological examination

78 of the mass, collected by ultrasound-guided puncture, suggested a malignant neoplasm of
79 mesenchymal origin.

80 Subsequently, due to the inability of the responsible party to perform an abdominal CT scan,
81 exploratory laparotomy and excision of the abdominal mass were chosen. Pre-operative
82 complementary examinations were performed, including blood count, serum biochemistry, chest X-
83 ray, abdominal ultrasound, electrocardiogram, and echocardiogram. The blood count revealed mild
84 anemia, lymphocytosis, and the presence of microfilariae. The echocardiogram revealed myxomatous
85 degeneration of the mitral valve, without hemodynamic repercussions. The electrocardiogram
86 showed an increased P wave, suggestive of left atrial overload. Radiographic examination revealed
87 enlargement of the right atrioventricular region, associated with a vascular pattern, suggesting heart
88 disease and pulmonary congestion. Preoperative abdominal ultrasound revealed, in addition to the
89 previously mentioned mass, bilateral nephropathy, mild gastropathy, dense biliary sludge, and
90 moderate splenomegaly. The ELISA test for *Dirofilaria immitis* was positive.

91 During an exploratory laparotomy, it was observed that the abdominal mass originated from
92 the cecum, and a typhlectomy was performed. The excised mass was transmural, with a multinodular
93 and irregular appearance, and measured approximately 7.0 cm in diameter. It presented a white and
94 dark brown color, a soft consistency, and, upon cutting, a smooth, solid, white and brown surface
95 (Fig. 1). The sample was fixed in 10% buffered formalin and subjected to routine histological
96 processing to obtain 3 to 5 μm sections, which were then stained with hematoxylin and eosin (2, 18).

97 Microscopic analysis revealed densely cellular, multinodular neoplastic proliferation in the
98 intestinal muscular layer, continuous with the mucosal layer, involving the intestinal crypts (Fig. 2A
99 and D). The cells were organized in a storiform arrangement, with solid areas, supported by a well-
100 vascularized, abundant fibrocollagenous stroma with areas of loose matrix. They appeared fusiform
101 to epithelioid (Fig. 2B), with eosinophilic cytoplasm, elongated or cigar-shaped nuclei, coarse
102 chromatin, and prominent nucleoli. There was marked anisocytosis and anisokaryosis, karyomegaly,
103 and frequent bizarre cells, in addition to eosinophilic intranuclear pseudoinclusions resulting from

104 cytoplasmic invagination (Fig. 2C). Extensive areas of hemorrhage were also observed, in addition
105 to lymphoplasmacytic inflammatory infiltrate in the mucosal region. Ten mitotic figures, both typical
106 and atypical, were counted in 15 fields (40x objective/FN18/2.37 mm²). The morphological diagnosis
107 was compatible with spindle cell sarcoma, suggestive of GIST or leiomyosarcoma.

108 For diagnostic confirmation, the material was submitted to immunohistochemical examination
109 with the markers CD117 (c-Kit) and Desmin. A primary mouse monoclonal antibody anti-c-Kit
110 (ThermoFisher, code 180066) was used at a 1:200 dilution and a mouse monoclonal antibody anti-
111 Desmin (Cell Marque, clone D33) at a 1:1000 dilution (12). In both cases, antigen retrieval was
112 performed in citrate solution pH 6, in a commercial pressure cooker (Muscae Plus, Erviegas). The
113 detection system used was Envision (HRP polymer, Dako), with diaminobenzidine (DAB) as
114 chromogen and counterstaining with hematoxylin (12). Protein blocking was performed with a
115 commercial universal solution (Dako), while endogenous peroxidase was inactivated with 8%
116 hydrogen peroxide in methanol (Dinâmica) (12). Canine pancreas was used as a positive control for
117 c-Kit and skeletal muscle for desmin, and mouse immunoglobulin (ThermoFisher) was replaced as a
118 negative control (12). Neoplastic cells exhibited diffuse and intense cytoplasmic immunostaining for
119 c-Kit, confirming the diagnosis of GIST (Fig. 3A-C), while the absence of desmin staining (Fig. 3D)
120 ruled out the hypothesis of leiomyosarcoma.

121

122 **Discussion**

123

124 Clinically, the signs observed in dogs with GISTs are usually nonspecific, such as vomiting,
125 diarrhea, anorexia, weight loss, and abdominal pain (1, 4, 15). This nonspecificity results from the
126 expansive growth of the mass, which promotes mechanical compression of adjacent intestinal loops,
127 altered motility, and partial obstruction of intestinal transit. Furthermore, ulceration of the overlying
128 mucosa can result in intraluminal hemorrhage, leading to melena or anemia (9, 15). In some cases,
129 clinical signs only manifest in advanced stages, when the tumor volume or ulceration becomes

130 clinically significant (15). In the present report, the presence of mild anemia may be associated with
131 both diagnosed dirofilariasis and micro-hemorrhages of the cecal mucosa and paraneoplastic
132 syndrome.

133 The diagnosis of gastrointestinal stromal tumor was established based on ultrasound,
134 histopathological, and immunohistochemical findings. The ultrasound revealed a mass in the
135 mesogastric region, without a clear definition of origin, a finding consistent with descriptions of
136 GISTs, which generally present as large, expansive masses that make it difficult to determine the
137 primary site until surgical exploration (16). In the present case, the cecal origin was only confirmed
138 during laparotomy, reinforcing the importance of intraoperative assessment for anatomical definition.
139 This finding corroborates reports that the cecum is one of the main sites of GIST involvement in dogs,
140 reinforcing the need for detailed inspection of this region in suspected cases (9, 13).

141 Microscopically, the neoplasm was predominantly composed of spindle-shaped cells in
142 storiform arrangement, with solid areas, characteristics more frequent in canine GISTs (1, 4).
143 However, polygonal cells with an epithelioid appearance were also observed, which, although less
144 common, are part of the morphological spectrum described in GISTs, and may occur in isolation or
145 mixed with the spindle pattern (9, 16). The coexistence of these patterns reinforces the morphological
146 heterogeneity of the neoplasm and the limitations of diagnosis based solely on conventional
147 histopathology. Intranuclear pseudoinclusions are cited in different neoplasms; however, in cases of
148 GISTs, they are correlated with greater cellular pleomorphism and malignancy (20, 21).

149 The simultaneous presence of fusiform and epithelioid patterns can lead to diagnostic
150 difficulties when considering only morphology, since smooth muscle neoplasms or even poorly
151 differentiated carcinomas can be considered differential diagnoses (19). In this context, diffuse
152 positive staining for c-Kit was crucial for confirming GIST, while the absence of Desmin expression
153 allowed for ruling out leiomyosarcoma. It is worth highlighting that, in cases of c-kit negativity, the
154 use of markers such as DOG1 can be fundamental for diagnostic confirmation, especially in cases
155 with a predominantly epithelioid pattern (5, 17).

156 The choice of canine pancreas as a positive control was appropriate, since this tissue exhibits
157 basal expression of c-Kit in some cell populations, ensuring the reactivity of the technique (7, 16).
158 This methodological care strengthens the reliability of the results, since the standardization of
159 immunohistochemistry is still a challenge in laboratories, and may influence diagnostic sensitivity.

160 GISTs frequently exhibit activating mutations in the c-KIT gene, generating direct implications
161 for therapeutic management, with the possibility of using tyrosine kinase inhibitors, such as toceranib
162 phosphate, in adjuvant therapy, which has demonstrated considerable prognostic efficacy (1, 6, 11).
163 In contrast, leiomyosarcomas generally do not present such mutations, making the use of conventional
164 cytotoxic chemotherapy, such as doxorubicin, preferable. Therefore, from a clinicopathological point
165 of view, confirmation of GIST is relevant because, in addition to characterizing the histogenetic origin
166 of the neoplasm, it guides potential therapeutic strategies. In the present case, both cell populations,
167 spindle-shaped and solid, showed positive immunostaining for c-Kit, confirming the homogeneity of
168 the immunophenotypic profile of the neoplasm, even in the face of morphological heterogeneity. This
169 result reinforces that despite histological variation, GIST maintains a consistent immunophenotypic
170 pattern, which facilitates its differentiation from other sarcomas.

171 Furthermore, recent studies have suggested that tumor size (>5 cm) and elevated mitotic index
172 are associated with a worse prognosis in dogs with GISTs, parameters that can be used in
173 complementary analyses and in future work for prognostic stratification. In the present case, the
174 findings were consistent with these negative prognostic indicators, as the specimen exceeded the 5
175 cm threshold and exhibited a high mitotic rate per high-power field. This correlation reinforces the
176 aggressive biological behavior observed and underscores the importance of such parameters in
177 predicting clinical outcomes for the patient. (5, 11).

178 Therefore, this case reinforces that the combination of morphological findings, even in the
179 presence of cellular heterogeneity (fusiform and epithelioid), with immunohistochemistry is essential
180 for the accurate diagnosis of GISTs in dogs, avoiding diagnostic errors and enabling more assertive
181 therapeutic decisions. The integration of imaging exams, detailed anatomopathological analysis, and

182 immunophenotyping should be considered the gold standard for the diagnosis of mesenchymal
183 gastrointestinal neoplasms in veterinary medicine.

184

185 **Data Availability**

186 All the original contributions presented in this study are included in the article/supplementary
187 material. Further inquiries can be directed to the corresponding author.

188

189 **Author Contributions**

190 **Lucas Costa Nogueira:** methodology, formal analysis, investigation, data curation, and writing
191 – original draft. **Virgilio Zoppi Lemos:** methodology, formal analysis, investigation, data curation,
192 and writing – review and editing. **Luise Klawa Jubrael:** methodology, formal analysis, investigation,
193 data curation, and writing – review and editing. **Tábata Maués:** methodology, formal analysis,
194 investigation, data curation, and writing – review and editing. **Angélica Consalter:** methodology,
195 formal analysis, investigation, data curation, and writing – review and editing. **Marcela Freire**
196 **Vallim de Mello:** methodology, formal analysis, investigation, data curation, and writing – review
197 and editing.

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199 **Conflict of Interest**

200 The authors declare no competing interests.

201

202 **Generative AI Use Statement**

203 The authors did not use generative artificial intelligence tools or technologies in creating or
204 editing any part of this manuscript.

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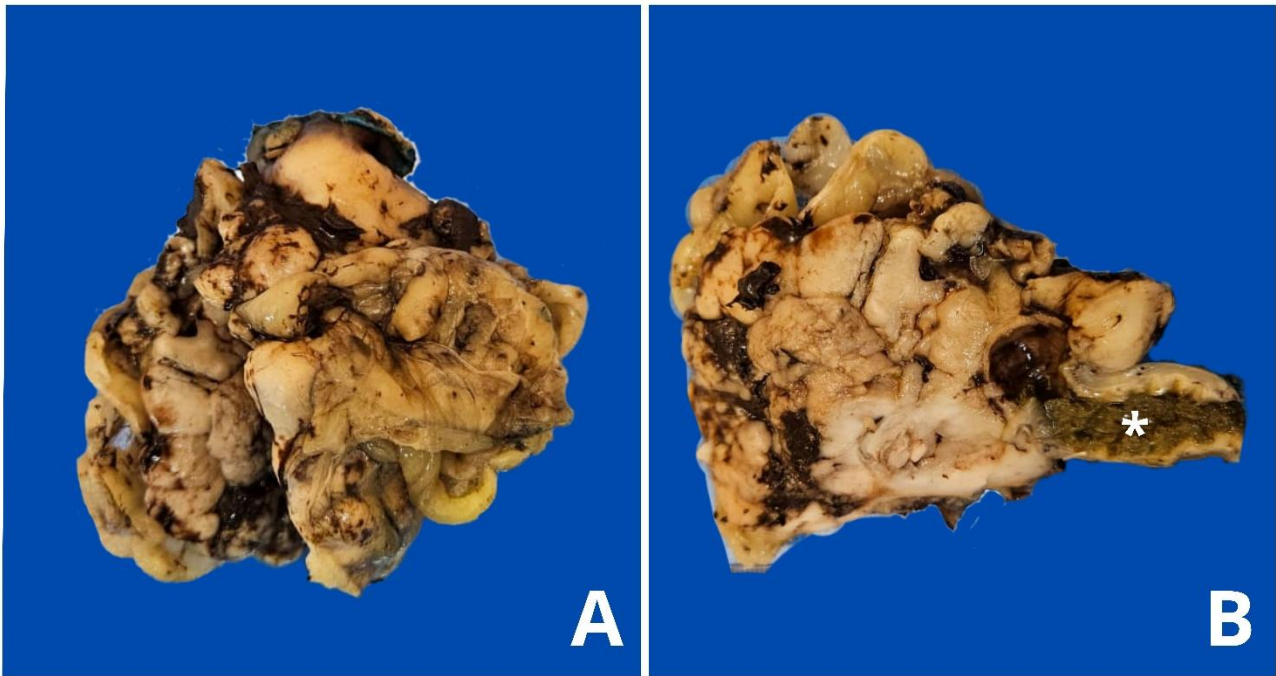
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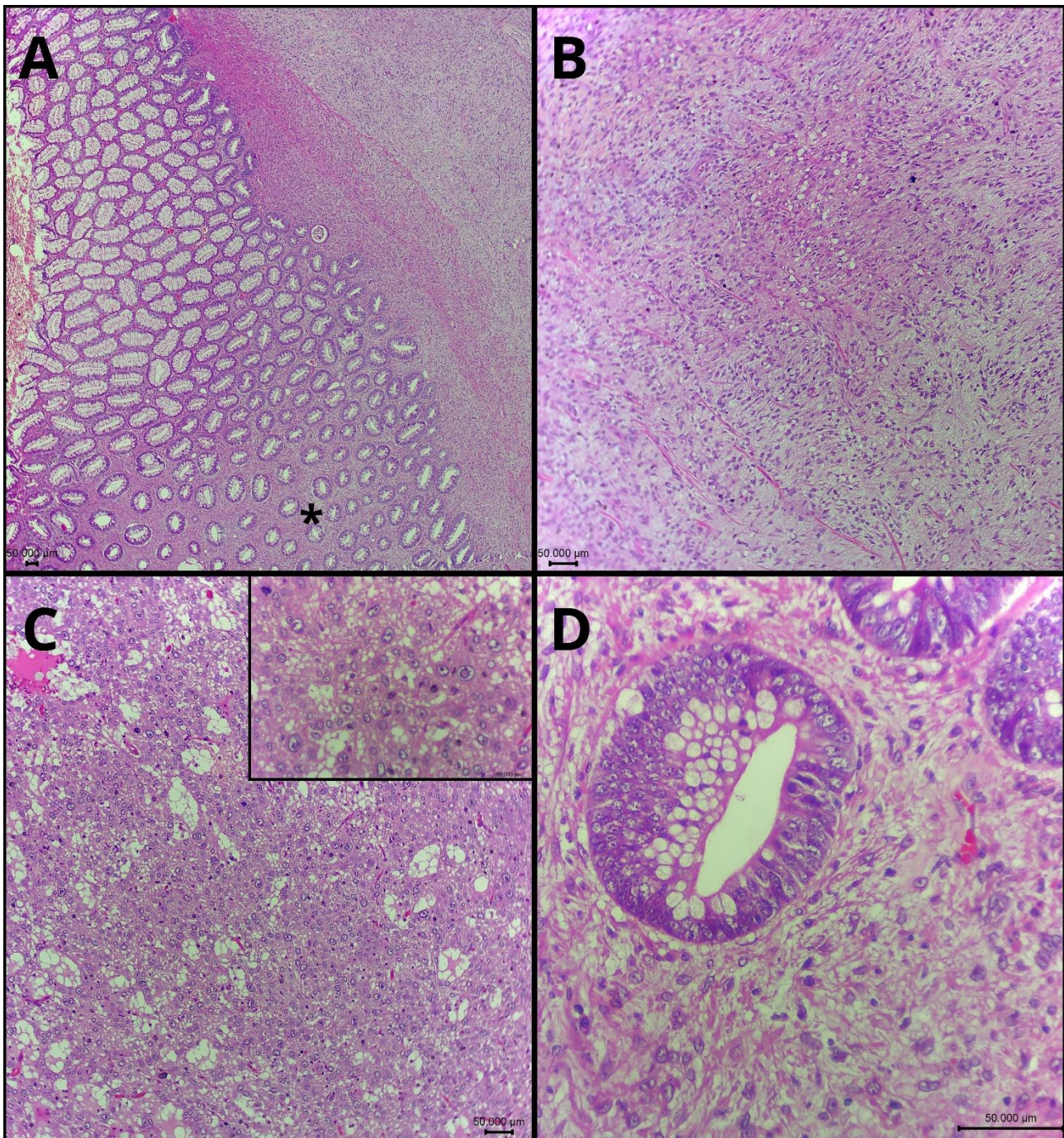
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273

274 **Figure 1.** Neoplastic mass in the cecum of a dog. A) Macroscopic aspect of the excised transmurular
275 mass, irregular, multinodular, 7.0 cm in diameter and soft consistency. B) Partial section showing the
276 intestinal lumen (*).

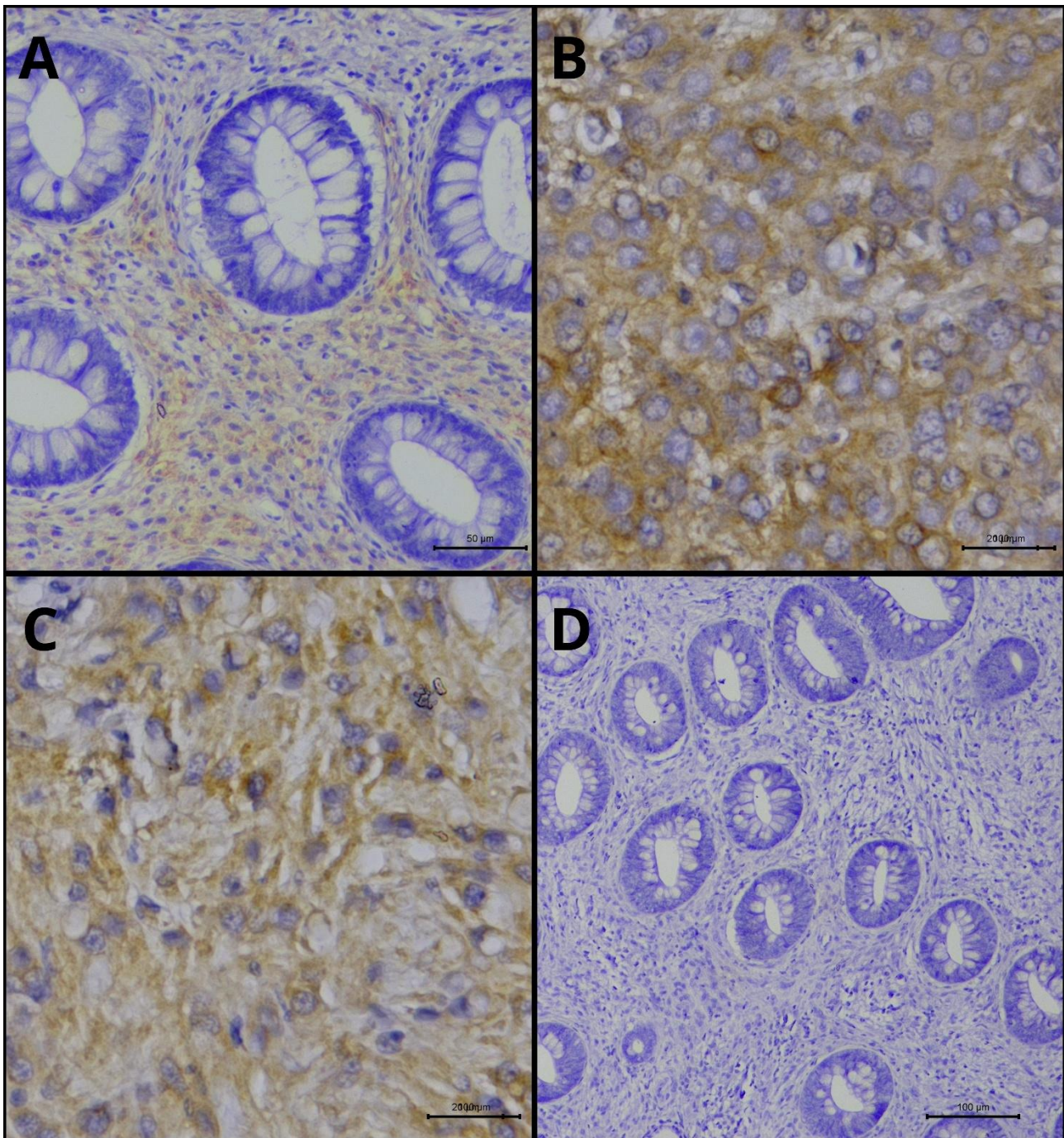
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279 **Figure 2.** Gastrointestinal stromal tumor, intestine, dog. A) Neoplastic proliferation in the muscular
 280 layer, dissecting the mucosa (arrow). Note the attenuation of the intestinal crypts interspersed with
 281 neoplastic bundles (*). B) Area composed of spindle cells. C) Area composed of epithelioid cells.
 282 Insert: Pleomorphic neoplastic cells, with mitotic figures and intranuclear pseudoinclusions (H&E,
 283 400X). D) Intestinal crypt surrounded by spindle-shaped neoplastic cells. Hematoxylin and eosin
 284 staining, A=40X, B and C=100X and D=400X.

285



286

287 **Figure 3.** Gastrointestinal stromal tumor, intestine, dog. Immunohistochemical examination. A)
288 Positive staining for c-kit in the plasma membrane of spindle cells surrounding the intestinal mucosal
289 crypts. B) Positive staining for c-kit in the plasma membrane of neoplastic polygonal cells. C) Positive
290 staining for c-kit in the plasma membrane of neoplastic spindle cells. D) Absence of staining for
291 desmin. DAB chromogen, haematoxylin counterstain.